

119b: METHANOMASSILIICOCCUS MEDIUM

| | | |
|--|--------|----|
| KH ₂ PO ₄ | 0.50 | g |
| MgSO ₄ x 7 H ₂ O | 0.40 | g |
| NaCl | 0.40 | g |
| NH ₄ Cl | 0.40 | g |
| CaCl ₂ x 2 H ₂ O | 0.05 | g |
| Trace element solution SL-10 | 1.00 | ml |
| Selenite-tungstate solution | 1.00 | ml |
| FeSO₄ x 7 H₂O solution (0.1% w/v) | 2.00 | ml |
| Yeast extract (OXOID) | 1.00 | g |
| Na-acetate | 1.00 | g |
| Na-formate | 2.00 | g |
| Sludge fluid | 50.00 | ml |
| Fatty acid mixture | 20.00 | ml |
| Sodium resazurin (0.1% w/v) | 0.50 | ml |
| Methanol (15% v/v) | 10.00 | ml |
| NaHCO ₃ | 1.00 | g |
| L-Cysteine HCl x H ₂ O | 0.50 | g |
| Na ₂ S x 9 H ₂ O | 0.50 | g |
| Distilled water | 940.00 | ml |

1. Dissolve ingredients except methanol, bicarbonate, cysteine and sulfide. Adjust pH of medium to 7.2 and sparge with 100% N₂ gas for 30 - 45 min to make it anoxic. Then dispense medium under same gas atmosphere into anoxic Hungate-type tubes to 30% v/v of their volume and autoclave. Add methanol (from 15% v/v stock solution), cysteine and sulfide from sterile anoxic stock solutions prepared under 100% N₂ gas and bicarbonate from a sterile anoxic stock solution prepared under 80% N₂ and 20% CO₂ gas mixture. Prior to use check pH of complete medium and adjust to 7.6, if necessary.
2. After inoculation, add sterile 80% H₂ and 20% CO₂ gas mixture to 0.5 bar overpressure. After growth becomes visible overpressure of 80% H₂ and 20% CO₂ gas mixture can be increased to 1 bar.
3. Note: Use 10% (v/v) as inoculum.

Trace element solution SL-10 (from medium 320)

| | | |
|--|--------|----|
| HCl (25%) | 10.00 | ml |
| FeCl ₂ x 4 H ₂ O | 1.50 | g |
| ZnCl ₂ | 70.00 | mg |
| MnCl ₂ x 4 H ₂ O | 100.00 | mg |
| H ₃ BO ₃ | 6.00 | mg |
| CoCl ₂ x 6 H ₂ O | 190.00 | mg |

119b: METHANOMASSILIICOCCUS MEDIUM

| | | |
|--|--------|----|
| $\text{CuCl}_2 \times 2 \text{ H}_2\text{O}$ | 2.00 | mg |
| $\text{NiCl}_2 \times 6 \text{ H}_2\text{O}$ | 24.00 | mg |
| $\text{Na}_2\text{MoO}_4 \times 2 \text{ H}_2\text{O}$ | 36.00 | mg |
| Distilled water | 990.00 | ml |

First dissolve FeCl_2 in the HCl, then dilute in water, add and dissolve the other salts. Finally make up to 1000.00 ml.

Selenite-tungstate solution (from medium 385)

| | | |
|--|---------|----|
| NaOH | 0.50 | g |
| $\text{Na}_2\text{SeO}_3 \times 5 \text{ H}_2\text{O}$ | 3.00 | mg |
| $\text{Na}_2\text{WO}_4 \times 2 \text{ H}_2\text{O}$ | 4.00 | mg |
| Distilled water | 1000.00 | ml |

Sludge fluid (from medium 119)

| | | |
|---------------|---------|----|
| Yeast extract | 4.00 | g |
| Sludge | 1000.00 | ml |

Add 0.4% yeast extract to sludge from an anaerobic digester, and after gassing with nitrogen gas for a few minutes incubate it at 37°C for 24 hours. Then centrifuge the sludge at 13000 g and autoclave the resulting, clear supernatant in screw-capped vessels under nitrogen gas. The sludge fluid can be stored at 8-12°C in the dark.

Fatty acid mixture (from medium 119)

| | | |
|-------------------------|--------|----|
| Isobutyric acid | 23.00 | ml |
| DL-2-Methylbutyric acid | 27.00 | ml |
| Valeric acid | 27.00 | ml |
| Isovaleric acid | 27.00 | ml |
| Distilled water | 896.00 | ml |

Adjust pH to 7.5 with concentrated NaOH.

$\text{FeSO}_4 \times 7 \text{ H}_2\text{O}$ solution (0.1% w/v) (from medium 119)

| | | |
|--|---------|----|
| $\text{FeSO}_4 \times 7 \text{ H}_2\text{O}$ | 1.00 | g |
| H_2SO_4 (0.1 N) | 1000.00 | ml |

The ferrous sulfate solution is not stable and should be freshly prepared.