Microorganisms



1514. Grace's Insect Medium

Medium for maintenance of SF-9 cells (ACC 125) for cultivation of Rhabdochlamydiae.

Grace's Insect Medium (Sigma Aldrich, G8142)	900	ml
Fetal Bovine Serum (heat-inactivated, 56°C, 30 min)	100	ml

Filter-sterilize

Maintenance conditions of SF-9 cells (see also DSMZ catalogue for ACC 125) SF-9 cells are cultivated at 25 – 29°C without the addition of CO_2 in flasks (25 cm²) with tightly closed lids. When a confluent layer has formed, infection can be carried out.

Infection with Rhabdochlamydiae

Exchange medium and add $500 - 1000 \,\mu l$ of EB stock solution (thawed quickly to 37° C). Centrifuge for 1 h onto the cell layer at 1600 rpm at 20° C.

Maintenance/Passage of infected insect cell cultures

Infected cultures can be maintained e.g. in 25 cm2 flasks (e.g. Nunc flasks, VWR, 734-2081) at 27° C in an incubator (no special requirements concerning CO_2 or light).

If the percentage of infected cells is still low (which might occur initially after inoculation of cultures with bacteria from a frozen stock) infected cultures may be maintained by weekly passage (1:5). Infection of yet uninfected cells by added bacteria or bacteria released from neighboring infected cells is facilitated by centrifugation (130 x g, 15 min) of the culture flask.

In well-infected cultures host cell lysis will be visible (the bacteria cause host cell lysis within approximately 5-7 days after invasion of host cells). To maintain such cultures, bacteria have to be transferred by adding culture supernatant (approximately 0.5-1 ml) to fresh culture flasks containing uninfected insect cells (approximately semi-confluent culture) each 7-10 days.

Storage

Bacteria released from host cells can be stored in

Sucrose-Phosphate-Glutamate	buffer
Cularana	

Sucrose	75	g
KH_2PO_4	0.52	g
$Na_2HPO_4 \times 2H_2O$	1.53	g
Glutamic acid	0.75	g
Distilled water	1000	ml

at -80°C for several months.

Infected cell cultures (supplemented with 10% DMSO) are best stored in liquid nitrogen.

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