

334a. METHANOTHRIX SOEHNGENII MEDIUM

Solution A:

KH ₂ PO ₄	0.22	g
Na ₂ HPO ₄ x 2 H ₂ O	0.86	g
Na-acetate	6.80	g
Na ₂ -EDTA solution (0.1% w/v)	1.00	ml
Trace elements solution (see medium 334)	10.00	ml
Na-resazurin solution (0.1% w/v)	0.50	ml
Cell-free culture supernatant of <i>S. associata</i> (see below)	100.00	ml
Distilled water	850.00	ml

Solution B:

Na ₂ CO ₃	1.25	g
Distilled water	25.00	ml

Solution C:

NaCl	0.30	g
CaCl ₂ x 2 H ₂ O	0.11	g
MgCl ₂ x 6 H ₂ O	0.10	g
Distilled water	10.00	ml

Solution D:

NH ₄ Cl	0.30	g
Distilled water	10.00	ml

Solution E:

Vitamins solution (see medium 141)	10.00	ml
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Solution F:

Na ₂ S x 9 H ₂ O	0.50	g
Distilled water	10.00	ml

Sparge *solution A* with 80% N₂ and 20% CO₂ gas mixture for 30 – 45 min to make it anoxic, dispense under same gas atmosphere into anoxic serum vials (e.g., 20 ml medium in 50 ml bottles) and autoclave. Prior to inoculation complete the medium by adding appropriate amounts of sterile *solutions C, D, E* and *F* prepared under 100% N₂ gas and *solution B* prepared under 80% N₂ and 20% CO₂ gas atmosphere. Vitamins are sterilized by filtration. Adjust the pH of the complete medium to 7.2.

Note: Use 20% (v/v) inoculum.

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Cell-free culture supernatant of Sphaerochaeta associata DSM 26261^T:

Cultivate DSM 26261^T in DSM medium 119 supplemented with 1.00 g/l glucose at 28°C until stationary phase is reached (7-10 days). Centrifuge culture at 3500 *xg* for 30 min, then sterilize supernatant by filtration and add it to the anoxic *solution A* before autoclaving.

Note: It is also possible to add 50.00 ml/l of a grown S. associata culture to solution A before autoclaving, provided bacterial DNA from dead cells is not a problem in the downstream processing of the Methanotherix culture.

For DSM 3870, DSM 4774 and DSM 6194 supplement medium with 10.00 ml/l of a coenzyme M (2-mercaptoethanesulfonate) solution (1.42% w/v, prepared under 100% N₂ gas and sterilized by filtration) and adjust pH of complete medium to 7.0.