

## 441. DIAZOTROPHIC MEDIUM (RBA)

**Solution A:**

$\text{KH}_2\text{PO}_4$	0.100	g
$\text{K}_2\text{HPO}_4$	0.900	g
NaCl	0.100	g
$\text{CaCl}_2 \times 2 \text{ H}_2\text{O}$	0.100	g
$\text{MgSO}_4 \times 7 \text{ H}_2\text{O}$	0.100	g
$\text{Na}_2\text{MoO}_4 \times 2 \text{ H}_2\text{O}$	0.005	g
$\text{NaVO}_3 \times \text{H}_2\text{O}$	0.005	g
$\text{MnSO}_4 \times \text{H}_2\text{O}$	0.005	g
$\text{FeSO}_4 \times 7 \text{ H}_2\text{O}$	0.010	g
Yeast extract	0.050	g
Trace element sol. SL-6 (see medium 27)	3.000	ml
Distilled water	950.000	ml
Agar (if necessary)	15.000	g

Adjust pH to 7.3.

**Solution B:**

Na <sub>2</sub> -succinate	1.000	g
DL-Malate	2.000	g
Na-pyruvate	1.000	g
D-Mannitol	2.000	g
D-Glucose	2.000	g
Distilled water	50.000	ml

Adjust pH to 7.3.

Sterilize solution A separately at 121°C for 15 min., cool to 50°C and then mix aseptically with filter-sterilized solution B and 5.0 ml of filter-sterilized standard vitamin solution (see medium 428).

RBA is an ammonium-free medium which has successfully been used for the isolation, growth and purity check of a broad spectrum of nitrogen fixing bacteria (Ref. 3363). For microaerophilic nitrogen-fixing bacteria use semisolid medium with 0.3% end concentration of agar and incubate the liquid cultures under 10% (v/v) air and 90% (v/v) N<sub>2</sub>.