

**853. FUSIBACTER PAUCIVORANS MEDIUM**

NH <sub>4</sub> Cl	1.00	g
K <sub>2</sub> HPO <sub>4</sub>	0.30	g
KH <sub>2</sub> PO <sub>4</sub>	0.30	g
MgCl <sub>2</sub> x 6 H <sub>2</sub> O	3.00	g
CaCl <sub>2</sub> x 2 H <sub>2</sub> O	0.10	g
KCl	1.00	g
NaCl	30.00	g
Yeast extract (OXOID)	1.00	g
Trypticase peptone (BD BBL)	1.00	g
Na-acetate x 3 H <sub>2</sub> O	0.50	g
Trace element solution (see medium 141)	10.00	ml
Na-resazurin solution (0.1% w/v)	0.50	ml
Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub> x 5 H <sub>2</sub> O	3.16	g
L-Cysteine-HCl x H <sub>2</sub> O	0.50	g
Na <sub>2</sub> CO <sub>3</sub>	1.50	g
D-Glucose	3.60	g
Na <sub>2</sub> S x 9 H <sub>2</sub> O	0.30	g
Distilled water	1000.00	ml

Dissolve ingredients, except thiosulfate, cysteine, carbonate, glucose and sulfide, then sparge medium for 30 - 45 min with 80% N<sub>2</sub> and 20% CO<sub>2</sub> gas mixture to make it anoxic. Add thiosulfate and cysteine, then adjust pH to 7.0 and dispense medium under 80% N<sub>2</sub> and 20% CO<sub>2</sub> gas atmosphere into anoxic Hungate-type tubes or serum vials and autoclave. After autoclaving, add glucose and sulfide from sterile anoxic stock solutions prepared under 100% N<sub>2</sub> gas and carbonate from a sterile anoxic stock solution prepared under 80% N<sub>2</sub> and 20% CO<sub>2</sub> gas atmosphere. Adjust pH of complete medium to 7.2 – 7.4, if necessary.