853b. FUSIBACTER BIZERTENSIS MEDIUM

NH_4Cl	1.00	g
K_2HPO_4	0.30	g
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$MgCl_2$ x 6 H_2O	0.50	g
$CaCl_2$ x 2 H_2O	0.10	g
KCl	0.10	g
NaCl	3.00	g
Yeast extract (OXOID)	1.00	g
Trypticase peptone (BD BBL)	1.00	g
Trace element solution (see medium 141)	10.00	ml
Na-resazurin solution (0.1% w/v)	0.50	ml
$Na_2S_2O_3$ x 5 H_2O	3.16	g
L-Cysteine-HCl x H_2O	0.50	g
$NaHCO_3$	4.00	g
D-Glucose	3.60	g
Na_2S x 9 H_2O	0.30	g
Distilled water	1000.00	ml

Dissolve ingredients, except thiosulfate, cysteine, bicarbonate, glucose and sulfide, then sparge medium for 30 - 45 min with 80% N_2 and 20% CO_2 gas mixture to make it anoxic. Add thiosulfate and cysteine, then adjust pH to 7.0 and dispense medium under 80% N_2 and 20% CO_2 gas atmosphere into anoxic Hungate-type tubes or serum vials and autoclave. After autoclaving, add glucose and sulfide from sterile anoxic stock solutions prepared under 100% N_2 gas and bicarbonate from a sterile anoxic stock solution prepared under 80% N_2 and 20% CO_2 gas atmosphere. Adjust pH of complete medium to 7.2 – 7.4, if necessary.