

**872. PAPILLIBACTER MEDIUM**

NH <sub>4</sub> Cl	0.30	g
K <sub>2</sub> HPO <sub>4</sub>	0.20	g
MgCl <sub>2</sub> x 6 H <sub>2</sub> O	0.40	g
CaCl <sub>2</sub> x 2 H <sub>2</sub> O	0.10	g
KCl	0.50	g
NaCl	1.00	g
Yeast extract (BD Bacto)	5.00	g
Trypticase peptone (BD BBL)	5.00	g
Trace element solution SL-10 (see medium 320)	1.00	ml
Na-resazurin solution (0.1% w/v)	0.50	ml
NaHCO <sub>3</sub>	4.50	g
<i>trans</i> -Cinnamic acid	0.75	g
L-Cysteine-HCl x H <sub>2</sub> O	0.50	g
Na <sub>2</sub> S x 9 H <sub>2</sub> O	0.50	g
Distilled water	1000.00	ml

Dissolve all ingredients except bicarbonate, cinnamic acid, sulfide and cysteine, then sparge medium with 80% N<sub>2</sub> and 20% CO<sub>2</sub> gas mixture for 30 – 45 min to make it anoxic. Dispense medium under same gas atmosphere into anoxic Hungate-type tubes or serum vials and autoclave. Add cinnamic acid, sulfide and cysteine from sterile anoxic stock solutions prepared under 100% N<sub>2</sub> gas and bicarbonate from a sterile anoxic stock solution prepared under 80% N<sub>2</sub> and 20% CO<sub>2</sub> gas atmosphere. Adjust pH of complete medium to 7.2 – 7.5, if necessary.