9. VY/2 AGAR

Baker's yeast	5.00	g
$CaCl_2$ x 2 H_2O	1.36	g
Vitamin B_{12}	0.50	mg
Agar (Difco)	15.00	g
Distilled water	1000.00	ml

Sterilize vitamin B_{12} separately by filtration. Prepare and store yeast cells as autoclaved stock suspension (5 g baker's yeast/100 ml distilled water, adjust pH to 6.5 and autoclave). Adjust pH of medium to 7.2 with KOH before, and after autoclaving and cooling to 50°C (use pH-indicator paper).

For DSM 11116 reduce amount of vitamin B_{12} to 0.05 mg/l.

Cultures of myxobacteria delivered freeze-dried:
Please see our video tutorial and follow the special instructions: 'Reactivation of Myxobacteria' given with the strain entry of our catalogue. For suspending the freeze-dried cells from ampoules, add about 0.5 - 1.0 ml medium MD1 (per liter: casiton 3.0 g; calciumchloride dihydrate 0.7 g; magnesiumsulphate heptahydrate 2.0 g) to the vial with freeze dried material.

Cultures of myxobacteria delivered as active cultures (growing on agar plates): Always use the rim of the swarm as inoculum for fresh media. If the swarms are creamy, transfer high amounts of cell mass to several spots on fresh VY/2 agar medium. If the swarm adheres to the agar or grows within the agar, cut small agar cubes from the rim of the swarm and place onto a fresh agar plate using an appropriate tool such as a lancet. Make sure that the pieces of swarm colonies grown on the agar are transferred to the agar plate. Attempt to place the inoculum in such a way that the swarms are in contact with the fresh agar plate.

Incubate for up to 3 weeks (in particular *Sorangium* and *Nannocystis* strains) at the temperature given for the strain, taking measures against desiccation. If there is no growth after ten days, carefully split up the agar-culture-cubes and squeeze the material to the agar plate and reincubate.