1011. SULFURIMONAS MJ MEDIUM

Dissolve ingredients (except bicarbonate, thiosulfate and vitamins), then sparge medium with 80% N₂ and 20% CO₂ gas mixture for 30 – 45 min to make it anoxic. Dispense medium under same gas atmosphere into anoxic Hungate-type tubes or serum vials up to a volume of 20% and autoclave. Add bicarbonate, thiosulfate and vitamins to the autoclaved medium from sterile anoxic stock solutions. Solutions of vitamins and thiosulfate are sterilized by filtration and stored under N₂, whereas the solution of bicarbonate is prepared under 80% N₂ and 20% CO₂ gas mixture and autoclaved. Adjust pH of complete medium to 6.7. After inoculation pressurize vessels to 0.5 bar overpressure with sterile 80% N₂ and 20% CO₂ gas mixture and add sterile air in an amount that is equivalent to a volume of 20% of the headspace.

For DSM 23290 supplement medium with 2.00 g/l NaNO₃. After inoculation pressurize vessels to 1 bar overpressure with sterile 80% N₂ and 20% CO₂ gas mixture. Do not add sterile air!

For DSM 24660 omit thiosulfate and supplement medium with 4.00 g/l yeast extract and 4.00 g/l Trypton peptone. After autoclaving the medium is reduced with 0.30 g/l Na₂S x 9 H₂O added from a sterile anoxic stock solution (3% w/v) prepared under 100% N₂ gas and the pH adjusted to 6.5. After inoculation pressurize vessels to 1 bar overpressure with sterile 80% N₂ and 20% CO₂ gas mixture. Do not add sterile air!

For DSM 28671 omit pressurizing vials with 80% N₂ and 20% CO₂ gas mixture.

Continued next page
For DSM 101688 and DSM 101780 supplement medium with 2.00 g/l KNO₃. Do not add overpressure of 80% N₂ and 20% CO₂ and do not add sterile air!