

## 1049. SELENATE REDUCER MEDIUM

### Solution A:

DL-Na-lactate	1.12	g
KCl	1.30	g
KH <sub>2</sub> PO <sub>4</sub>	0.20	g
NaCl	23.00	g
NH <sub>4</sub> Cl	0.50	g
CaCl <sub>2</sub> x 2 H <sub>2</sub> O	0.10	g
MgCl <sub>2</sub> x 6 H <sub>2</sub> O	3.00	g
Resazurin	0.50	mg
Agar	15.00	g
Distilled water	940.00	ml

### Solution B:

Na <sub>2</sub> SeO <sub>4</sub>	1.89	g
Distilled water	10.00	ml

### Solution C:

NaHCO <sub>3</sub>	1.00	g
Distilled water	10.00	ml

### Solution D:

Na <sub>2</sub> S x 9 H <sub>2</sub> O	0.10	g
Distilled water	10.00	ml

### Solution E:

Na <sub>2</sub> SeO <sub>3</sub>	0.003	g
Na <sub>2</sub> WO <sub>4</sub>	0.008	g
Distilled water	100.00	ml

### Solution F:

Vitamin solution (see medium 461)

### Solution G:

Trace element solution SL-10 (see medium 320)	1.00	ml
Distilled water	9.00	ml

### Solution H:

1,4-Naphthoquinone	2.00	mg
Hemin	0.50	mg
Distilled water	100.00	ml

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After cooling of medium A, add per 100 ml:

- 1.0 ml solution B,
  - 2.5 ml solution C,
  - 1.0 ml solution D,
  - 0.1 ml solution E,
  - 0.5 ml solution F,
  - 1.0 ml solution G and
  - 1.0 ml solution H.
- Adjust pH to 7.0.

It is recommended to cultivate strain DSM 17993 on agar plates which may be prepared under aerobic conditions. Plates are then preincubated under anaerobic conditions prior to inoculation. They will stay pink coloured.

For liquid medium, prepare solution A, distribute to anaerobic cultivation vessels, heat to the boil and cool to room temperature while gassing with  $N_2 : CO_2$  (80:20). Close vessels and autoclave. Prepare solutions B, C, D, E and H under anaerobic conditions and autoclave. Solutions F and H are filter sterilized and flushed with  $N_2$ . Selenate (solution B) might be replaced by nitrate (solution: 1.00 g  $KNO_3$  in 10 ml).