

## 1095. NATRANAEROBIUS MEDIUM

NaCl	100.0	g
KH <sub>2</sub> PO <sub>4</sub>	0.2	g
MgCl <sub>2</sub>	0.1	g
NH <sub>4</sub> Cl	0.5	g
Yeast extract (OXOID)	10.0	g
Tryptone (BD Bacto)	10.0	g
Trace element solution SL-10 (see medium 320)	1.0	ml
Na <sub>2</sub> CO <sub>3</sub>	68.0	g
NaHCO <sub>3</sub>	38.0	g
L-Cysteine-HCl x H <sub>2</sub> O	0.7	g
Vitamin solution (see medium 141)	10.0	ml
Sucrose	5.0	g
Distilled water	1000.0	ml

Dissolve ingredients (except carbonates, cysteine, vitamins and sucrose), then sparge medium with 100% N<sub>2</sub> gas for 30 – 45 min to make it anoxic. Add carbonate, hydrogencarbonate and cysteine and adjust pH to 8.5. Dispense medium under 100% N<sub>2</sub> gas atmosphere into anoxic Hungate-type tubes or serum vials and autoclave. After autoclaving add vitamins and sucrose from sterile anoxic stock solutions prepared under 100% N<sub>2</sub> gas and sterilized by filtration.

For [DSM 18760](#) adjust pH of medium to 9.5.

For [DSM 18770](#) adjust pH of medium to 9.7 and replace sucrose with 5.0 g/l Casamino acids added from a sterile anoxic stock solution after autoclaving.

For [DSM 23380](#) adjust pH of medium to 9.5 and replace sucrose with 5.0 g/l D-glucose added from a sterile anoxic stock solution after autoclaving.