

**1163a. OCEANOTOGA MEDIUM**

|   |         |    |
|---|---------|----|
| NaCl                                    | 30.00   | g  |
| HEPES (SIGMA)                           | 4.70    | g  |
| KCl                                     | 0.33    | g  |
| MgCl <sub>2</sub> x 2 H <sub>2</sub> O  | 0.90    | g  |
| MgSO <sub>4</sub> x 7 H <sub>2</sub> O  | 1.40    | g  |
| NH <sub>4</sub> Cl                      | 0.25    | g  |
| CaCl <sub>2</sub> x 2 H <sub>2</sub> O  | 0.14    | g  |
| KH <sub>2</sub> PO <sub>4</sub>         | 0.45    | g  |
| Yeast extract                           | 5.00    | g  |
| Trace element solution (see medium 141) | 10.00   | ml |
| Na-resazurin solution (0.1% w/v)        | 0.50    | ml |
| L-Cysteine-HCl x H <sub>2</sub> O       | 0.50    | g  |
| D-Glucose                               | 5.00    | g  |
| Vitamin solution (see medium 141)       | 10.00   | ml |
| Distilled water                         | 1000.00 | ml |

Dissolve ingredients except cysteine, maltose and vitamins. Sparge medium with 100% N<sub>2</sub> gas for 30 – 45 min to make it anoxic, then add and dissolve cysteine and adjust pH to 7.5 with NaOH. Dispense medium under 100% N<sub>2</sub> gas atmosphere into anoxic Hungate-type tubes or serum vials and autoclave. Add glucose and vitamins after autoclaving from anoxic stock solutions prepared under 100% N<sub>2</sub> gas and sterilized by filtration.

For [DSM 24739](#) replace glucose with 5.00 g/l D-xylose and supplement medium after autoclaving with 2.50 g/l Na-thiosulfate added from an anoxic sterile stock solution.