

212: SYNTHROPHOMONAS MEDIUM

Solution A	942.00	ml
Solution B	30.00	ml
Solution C	10.00	ml
Solution D	1.00	ml
Solution E	10.00	ml
Solution F	10.00	ml

1. Add and dissolve ingredients of solution A and sparge medium with 80% N₂ and 20% CO₂ gas mixture for 30 - 45 min to make it anoxic. Dispense medium under the same gas atmosphere into anoxic Hungate-type tubes and autoclave. Solution B is prepared under 80% N₂ and 20% CO₂ gas atmosphere and autoclaved. Solutions C, E and F are autoclaved under 100% N₂ gas atmosphere. Solution D is prepared under 100% N₂ gas and sterilized by filtration. To complete the medium appropriate amounts of solutions B to F are added to the sterile solution A in the sequence as indicated. Adjust pH of complete medium to 7.2, if necessary.

2. Note: Some cultures are shipped in semi-solid medium which stimulates growth at the beginning. For agar stabs 3.00 g/l agar are added to the complete medium from a sterile anoxic stock solution (2% w/v). Upon receipt add anoxically 1 - 2 ml of the recommended freshly prepared liquid medium to the agar tube and incubate for 3 - 5 days . After incubation transfer 0.5 ml of the resulting cell suspension in the liquid phase to tubes with liquid medium.

For [DSM 2805](#): Replace butyrate with 1.50 g/l Na-propionate and supplement medium with 1.00 g/l D-glucose added from a sterile anoxic stock solution sterilized by filtration.

For [DSM 4212](#): Replace Na-butyrate in solution C with 0.61 g Na-stearate. Adjust the amount of distilled water in solution C to 20 ml. Dissolve stearate in water by heating in a water bath.

For [DSM 16706](#): Replace butyrate with 1.50 g/l Na-propionate.

For [DSM 26217](#): Omit butyrate and add to the complete medium 7.00 g/l sucrose from a sterile anoxic stock solution.

For [DSM 102352](#): Replace butyrate with 2.00 g/l Na-benzoate.

Solution A

KH ₂ PO ₄	0.50	g
MgCl ₂ x 6 H ₂ O	0.33	g
NaCl	0.40	g
NH ₄ Cl	0.40	g
CaCl ₂ x 2 H ₂ O	0.05	g
Trace element solution SL-10	1.00	ml

212: SYNTHROPHOMONAS MEDIUM

Selenite-tungstate solution	1.00	ml
Clarified rumen fluid	50.00	ml
Trypticase peptone (BD BBL)	1.00	g
Na ₂ SO ₄	2.80	g
Sodium resazurin (0.1% w/v)	0.50	ml
Distilled water	890.00	ml

Solution B

Na ₂ CO ₃	1.50	g
Distilled water	30.00	ml

Solution C

Na-butyrate	1.70	g
Distilled water	10.00	ml

Solution D (from medium 212)

Seven vitamins solution	1.00	ml
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Solution E (from medium 212)

L-Cysteine HCl x H ₂ O	0.30	g
Distilled water	10.00	ml

Solution F (from medium 212)

Na ₂ S x 9 H ₂ O	0.30	g
Distilled water	10.00	ml

Trace element solution SL-10 (from medium 320)

HCl (25%)	10.00	ml
FeCl ₂ x 4 H ₂ O	1.50	g
ZnCl ₂	70.00	mg
MnCl ₂ x 4 H ₂ O	100.00	mg
H ₃ BO ₃	6.00	mg
CoCl ₂ x 6 H ₂ O	190.00	mg
CuCl ₂ x 2 H ₂ O	2.00	mg
NiCl ₂ x 6 H ₂ O	24.00	mg
Na ₂ MoO ₄ x 2 H ₂ O	36.00	mg
Distilled water	990.00	ml

First dissolve FeCl₂ in the HCl, then dilute in water, add and dissolve the other salts. Finally make up to 1000.00 ml.

212: SYNTHROPHOMONAS MEDIUM

Selenite-tungstate solution (from medium 385)

NaOH	0.50	g
Na ₂ SeO ₃ x 5 H ₂ O	3.00	mg
Na ₂ WO ₄ x 2 H ₂ O	4.00	mg
Distilled water	1000.00	ml

Clarified rumen fluid (from medium 1310)

Rumen fluid from cow or sheep (obtained from fistulated animals or abattoir refuse) is filtered through muslin, autoclaved at 121°C for 15 min and then centrifuged at 27,000 g for 20 min. The supernatant is made anoxic by sparging with 100% N₂ gas for 15 min, dispensed under same gas atmosphere into anoxic serum vials to 30% of volume and then stored frozen at -20°C.

Seven vitamins solution (from medium 503)

Vitamin B ₁₂	100.00	mg
p-Aminobenzoic acid	80.00	mg
D-(+)-biotin	20.00	mg
Nicotinic acid	200.00	mg
Calcium pantothenate	100.00	mg
Pyridoxine hydrochloride	300.00	mg
Thiamine-HCl x 2 H ₂ O	200.00	mg
Distilled water	1000.00	ml