

## 212a. TREPONEMA RUMINIS MEDIUM

### Solution A:

KH <sub>2</sub> PO <sub>4</sub>	0.50	g
MgCl <sub>2</sub> x 6 H <sub>2</sub> O	0.33	g
NaCl	0.40	g
NH <sub>4</sub> Cl	0.40	g
CaCl <sub>2</sub> x 2 H <sub>2</sub> O	0.05	g
Trace element solution SL-10 (see medium 320)	1.00	ml
Selenite-tungstate solution (see medium 385)	1.00	ml
Rumen fluid, clarified (see medium 1310)	50.00	ml
Trypticase peptone (BD BBL)	1.00	g
Na <sub>2</sub> SO <sub>4</sub>	2.80	g
Vitamin K <sub>1</sub> solution (see medium 78)	1.00	ml
Volatile fatty acid mixture (see medium 330)	3.10	ml
Na-resazurin solution (0.1% w/v)	0.50	ml
Distilled water	860.00	ml

### Solution B:

Na <sub>2</sub> CO <sub>3</sub>	1.50	g
Distilled water	30.00	ml

### Solution C:

Soluble starch	0.40	g
Distilled water	4.00	ml

### Solution D:

D-Glucose	0.80	g
Distilled water	4.00	ml

### Solution E:

Sugar mix (see medium 843)	30.00	ml
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### Solution F:

Vitamin solution (see medium 503)	1.00	ml
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### Solution G:

Na-pyruvate	0.50	g
Distilled water	2.00	ml

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**Solution H:**

L-Cysteine-HCl x H <sub>2</sub> O	0.30	g
Distilled water	10.00	ml

**Solution I:**

DL-Dithiothreitol (DTT)	0.40	g
Distilled water	10.00	ml

Add and dissolve ingredients of *solution A*, adjust pH to 7.0 and sparge medium with 80% N<sub>2</sub> and 20% CO<sub>2</sub> gas mixture for 30 – 45 min to make it anoxic. Dispense medium under the same gas atmosphere into anoxic Hungate-type tubes and autoclave. *Solution B* is prepared under 80% N<sub>2</sub> and 20% CO<sub>2</sub> gas atmosphere and autoclaved. *Solutions C, D and H* are autoclaved under 100% N<sub>2</sub> gas atmosphere. *Solutions E, F, G and I* are prepared under 100% N<sub>2</sub> gas and sterilized by filtration. To complete the medium appropriate amounts of *solutions B to I* are added to the sterile *solution A* in the sequence as indicated.

*Note: Some cultures are shipped in semi-solid medium which stimulates growth at the beginning. For agar stabs 3.00 g/l agar are added to the complete medium from a sterile anoxic stock solution (2% w/v). Upon receipt add anoxically 1 - 2 ml of the recommended freshly prepared liquid medium to the agar tube and incubate for 3 - 5 days. After incubation transfer 0.5 ml of the resulting cell suspension in the liquid phase to tubes with liquid medium.*