

## 719. ANAEROFILUM MEDIUM

KH <sub>2</sub> PO <sub>4</sub>	0.50	g
MgSO <sub>4</sub> x 7 H <sub>2</sub> O	0.40	g
NaCl	0.40	g
NH <sub>4</sub> Cl	0.40	g
CaCl <sub>2</sub> x 2 H <sub>2</sub> O	0.05	g
FeSO <sub>4</sub> x 7 H <sub>2</sub> O solution (0.1% w/v in 0.1 N H <sub>2</sub> SO <sub>4</sub> )	2.00	ml
Trace element solution SL-10 (see medium 320)	1.00	ml
Yeast extract	1.00	g
Na-acetate	1.00	g
Na-formate	2.00	g
Fatty acid mixture (see medium 119)	20.00	ml
Na-resazurin solution (0.1% w/v)	0.50	ml
NaHCO <sub>3</sub>	4.00	g
D-Glucose	5.00	g
L-Cysteine-HCl x H <sub>2</sub> O	0.50	g
Na <sub>2</sub> S x 9 H <sub>2</sub> O	0.50	g
Distilled water	930.00	ml

Dissolve ingredients except bicarbonate, glucose, cysteine and sulfide. Sparge medium with 80% N<sub>2</sub> and 20% CO<sub>2</sub> gas mixture for 30 – 45 min to make it anoxic. Add and dissolve bicarbonate, then dispense medium under same gas atmosphere into anoxic Hungate-type tubes or serum vials and autoclave. Add glucose, cysteine and sulfide from sterile anoxic stock solutions prepared under 100% N<sub>2</sub> gas. Prior to use check pH of complete medium and adjust to 6.8 - 7.0, if necessary.