

830: R2A MEDIUM

Yeast extract	0.50	g
Proteose peptone (Difco no. 3)	0.50	g
Casamino acids	0.50	g
Glucose	0.50	g
Starch (soluble)	0.50	g
Na-pyruvate	0.30	g
K ₂ HPO ₄	0.30	g
$MgSO_4 \times 7 H_2O$	0.05	g
Agar	15.00	g
Distilled water	1000.00	ml

Final pH 7.2; adjust with crystalline K_2HPO_4 or KH_2PO_4 before adding agar. Add agar, heat medium to boiling to dissolve agar, and autoclave for 15 min at 121°C.

For <u>DSM 2406</u>: Rehydrate and grow freeze-dried cells from ampoule in liquid medium 54 or 830 first. Subsequent subcultivation may be carried out in liquid medium or on agar plates.

For DSM 4136: add 1% NaCl

For <u>DSM 6696</u>: Rehydate and grow freeze dried cells from ampoule in liquid medium 830. Subsequent subcultivation may be carried out in liquid medium or on agar plates, or in medium 457 with styrol

For <u>DSM 6708</u>: Rehydrate and grow lyophylisate in liquid medium 830. Subsequently, subculturing may be carried out in liquid medium or on agar plates, on medium 830, 464 or 465 as indicated

For <u>DSM 6780</u>: Reactivate freeze dried culture in medium 830 (liquid medium and agar plate) and incubate for up to 14 days

For <u>DSM 11199</u>: Aerobic; reactivate lyophilized cells in liquid medium 830, incubate for at least 10 days

For DSM 12184: liquid

For <u>DSM 12445</u>: Rehydrate and grow freeze-dried cells from the ampoule in medium 830, subsequent subculturing may be carried out in medium 457 with carbon source or in medium 830 or 464.

For <u>DSM 13665</u>: With or without 10 mg/l 2,4,6-trichlorophenol and 20 mg/l pentachlorophenol

For <u>DSM 14655</u>: Rehydrate and grow lyophilized cells from the ampoule in 5 ml liquid broth 1. Subsequent subculturing may be carried out in liquid medium or on agar plates

For <u>DSM 14692</u>: Aerobic, rehydrate and grow lyophilized cells in liquid medium. Subsequent subculturing may be carried out on agar plates also.

For DSM 14804: Only one passage beginning with mineral medium with hexane is possible

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For <u>DSM 14805</u>: Aerobic; reactivate freeze dried cells in liquid medium.

For DSM 15083: Add 1% methanol; aerobe

For <u>DSM 15124</u>: Aerobic. Rehydrate and grow lyophilized cells from ampoules in liquid broth 830. Subsequent subculturing may be carried out in liquid medium or on agar plates.

For DSM 15892: Cultivate in liquid medium, only thin colonies on agar plate

For <u>DSM 16361</u>: Omit Agar. Adjust pH of final medium to 6.0.

For <u>DSM 16616</u>: Rehydrate and grow lyophilized cells from the ampoule in 5 ml liquid broth. Subsequent subculturing may be carried out in liquid medium or on agar plates.

For <u>DSM 16845</u>: Rehydrate and grow freeze dried cells from ampoule in liquid medium 830. Subsequent subsculturing may be carried out in medium 465 with TM or on agar plates, too.

For <u>DSM 16968</u>: Plus MnSO₄ (10mg/l)

For DSM 16969, DSM 18135, DSM 21316, DSM 21346, DSM 21568: Plus MnSO₄

For <u>DSM 16976</u>, <u>DSM 29786</u>, <u>DSM 29790</u>, <u>DSM 29794</u>: PH 9.0

For <u>DSM 17513</u>: Rehydrate and grow freeze-dried cells from ampoule in liquid medium 830 first. Subsequent subcultivation may be carried out in liquid medium or on agar plates and in medium 457 with 3-chlorosalicylate.

For <u>DSM 17870</u>: Rehydrate and grow lyophilized cells from the ampoule in 5 ml liquid broth 830. Subsequent subculturing may be carried out in liquid medium or on agar plates

For <u>DSM 17871</u>: Plus MnSO₄ (10 mg/l)

For DSM 17997: Adjust pH to 10 by adding 200 ml 0,5M Na₂CO₃

For DSM 17998: Adjust pH to 10 by adding 200ml 0.5M sodium carbonate [Na₂CO₃]

For <u>DSM 17999</u>: Add 0.5 M Na₂CO₃, pH 10.0

For <u>DSM 18042</u>, <u>DSM 18043</u>, <u>DSM 18044</u>, <u>DSM 18045</u>, <u>DSM 45021</u>: Plus 10% NaCl

For DSM 18094, DSM 19250: Preferably liquid medium

For DSM 18203: Plus 2g/l methanol

For <u>DSM 18221</u>: Keep agar surface moist with a film of liquid medium

For <u>DSM 18504</u>, <u>DSM 18629</u>: adjust pH to 5.5

For DSM 18603: 10-fold diluted and supplemented with 0.05% glucose

For DSM 18644: Growth is better on solid media if the surface is covered with a layer of

liquid medium

For DSM 18693: Dilute the medium 10 fold and adjust the pH to 5.0-5.5

For DSM 18704: Dilute the medium 10 fold and adjust the pH to 4.5-5.0

For DSM 18758: Dilute the medium 1:10, adjust the pH to 5.5

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For DSM 18855: Microaerobic (candle jar)

For <u>DSM 19016</u>, <u>DSM 19017</u>, <u>DSM 19018</u>: Reactive in M. 1712; aerobic

For DSM 19019: Reactivate in M. 1712; aerobic

For DSM 19020: Reactive in M. 1712; aerob

For DSM 19021: Microaerobic, growing at 2-4% oxygen in atmosphere

For DSM 19096: (one-fifth-strength)

For DSM 19249: Preferably liquid medium

For DSM 19309: (dilute 1:5)

For <u>DSM 19717</u>, <u>DSM 19724</u>: With 5 g/l NaCl

For <u>DSM 19937</u>: PH: 9.5

For DSM 21036: Dilute medium 1:1 with distilled water

For DSM 21240: With NaCl (20g/l) and MgCl2x 6H2O (3g/l)

For DSM 21362: 1 drop of 10x Wolin Vitamine Solution (see medium 141) and 10 microliter

trace element solution SL-10 (see medium 320) for better growth in liquid

For <u>DSM 21439</u>: 1:2 diluted

For DSM 21490: 1:5 diluted, with cysteine and pyruvate (0.2 g/l each); keep agar surface

moist

For DSM 21594: At 20 °C or 28 °C; (this medium is recommended for the routine

cultivation of the strain)

For DSM 21599, DSM 21701: diluted 1:1

For DSM 21636: At half concentration

For DSM 21646: Keep agar surface moist and incubation atmosphere humid

For DSM 21648: Slowly growing, keep agar surface moist

For DSM 21670: With 10 mg/L MnSO₄, adjust to pH 9 with Na-sesquicarbonate

For <u>DSM 21675</u>: Adjust pH to 7.5

For DSM 21697: PH 7.2

For DSM 21712: Diluted 1:2

For <u>DSM 21788</u>, <u>DSM 21937</u>, <u>DSM 21938</u>, <u>DSM 23562</u>: PH 7

For DSM 21791: PH 6.2

For <u>DSM 21895</u>: Keep agar surface moist during the whole incubation period

For <u>DSM 21942</u>: Plus Na-sesquicarbonate solution (see medium 31); pH 9.0

For DSM 21953, DSM 21954: half concentrated and prepared with 75% seawater

For DSM 21994: Dilute R2A medium 1:5 and add Na-pyruvate and cysteine-HCl x H_2O , 2 g

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per L each

For DSM 22177, DSM 23415: half strengh

For <u>DSM 22274</u>, <u>DSM 22275</u>, <u>DSM 22383</u>, <u>DSM 22384</u>, <u>DSM 22899</u>, <u>DSM 22900</u>, <u>DSM</u>

23227, DSM 23324, DSM 23450, DSM 23456, DSM 23459: PH 7.0

For DSM 22342: Plus 10 mg/l MnSO₄

For <u>DSM 22349</u>: Diluted 1:5. Add Na-pyruvate and cysteine-HCl x H₂O, 0,2 g per L each

For DSM 22385: PH 6.0 - 8.0

For <u>DSM 22386</u>: PH 6.5 - 8.0

For DSM 22405: Plus 10 mg/L MnSO₄

For DSM 22464: Dilute the medium 20 fold and add 0.05% fructose, adjust pH 4.0 - 5.0

For DSM 22489: 20-fold diluted and supplied with 0.05% glucose or fructose, pH 5.0 - 5.8

For DSM 22513: PH 7.3

For DSM 22869: Plus 10 µg/ml MnSO₄

For DSM 23054: PH 7.0 - 8.0

For DSM 23119: Adjust the pH of the medium to 6.0

For <u>DSM 23128</u>, <u>DSM 23136</u>, <u>DSM 23137</u>, <u>DSM 23138</u>: Adjust pH to 5.0 - 5.6 with HCl.

Solidify with 5g/l agar

For <u>DSM 23228</u>: Plus 10 mg/L MnSO₄; pH 8.5

For <u>DSM 23480</u>: Half strength substrate concentration

For DSM 23489, DSM 23491: Plus 10 mg/L MnSO₄; pH 7.0

For <u>DSM 23490</u>: Plus 3% NaCl plus 10 mg/L MnSO₄; pH 7.0

For <u>DSM 23630</u>: (leave out phosphate and pyruvate. Dilute 1:10 and adjust pH to 4.5 - 5.0 with alginic acid (or MES)); fill tubes for apparately (Hungate) half with medium, class

with alginic acid (or MES)); fill tubes for anaerobes (Hungate) half with medium, close

tightly. Do not shake.

For DSM 23641: PH 5.5, use gelrite instead of agar

For DSM 23886: Dilute the medium 10 fold, add 0.02% glucose and adjust the pH to 5.0 -

5.5

For <u>DSM 23906</u>: Plus 0.1% Tween 80

For DSM 24297: 1:10 diluted, pH 4.5, without pyruvate, without phosphate

For DSM 24448: With 1mg/l riboflavine

For DSM 24462: Plus 100 µl Anaerobiersalze (salt solution of DSMZ Medium 104)

For <u>DSM 25158</u>: Subculturing is recommended to be carried out on agar plates because

growth in liquid medium is weak

For <u>DSM 25262</u>: Plus 1.0 g/l NaCl

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For <u>DSM 25320</u>: Plus 0,1% Tween 80

For DSM 25321: Plus Tween 80

For DSM 25461: With 5 ml vitamin solution, see e.g. medium 461

For DSM 26743: plus 0.5 g/l mannitol

For DSM 26776: PH 6.0

For DSM 27573: Keep agar surface moist during incubation. No visible growth in liquid

media

For DSM 27864: Plus vitamin solution (2ml/L); see medium 141

For DSM 27942: Liquid media require heavy inoculum

For DSM 28057, DSM 29854: PH 8.0

For DSM 28206: pH 5.5

For DSM 28215: pH 5,5

For <u>DSM 28344</u>: Rehydrate and grow lyophilized cells from the ampoule in 5 ml liquid broth. Subsequent subculturing may be carried out in liquid medium or on agar plates

For DSM 29249: 20% R2A; 20-37°C, pH 6.0-8.0 optimum

For DSM 29289: 1:10 diluted

For DSM 29414, DSM 45618, DSM 29840: PH 5.0

For DSM 45540: pH 5,3

For DSM 45816: pH 6.5

For DSM 100059: Only 25% strenth of the original medium 830

For <u>DSM 100467</u>, <u>DSM 100468</u>, <u>DSM 100482</u>: plus 30 g/l NaCl

For <u>DSM 100496</u>, <u>DSM 100572</u>: Plus 30 g/l NaCl

For DSM 100788: PH=9.0

For DSM 102969: Plus 1% methanol

For DSM 103219: add 0.1% methylammoniumchloride

For DSM 103337: Slow growth, poor growth in liquid media

For DSM 103476: Twofold concentrated; incubation period 6-8 d

For <u>DSM 104092</u>, <u>DSM 104093</u>: Add 0.06% sodium pyruvate

For DSM 105494: Plus 0,1% Methylamine

For DSM 105890: One-tenth-strength

For <u>DSM 106290</u>, <u>DSM 103404</u>, <u>DSM 106523</u>: pH 10, plus 2% NaCl

For DSM 106492: PH 6.6

For <u>DSM 106737</u>, <u>DSM 106674</u>, <u>DSM 106675</u>, <u>DSM 108426</u>: PH 8.5

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For DSM 108208: braucht pH 8.5, wächst nicht auf pH 7.2

For <u>DSM 108248</u>: 1/10 diluted

For DSM 108317: braucht pH 8.5, wächst nicht auf pH 7.2 nach LN

For <u>DSM 108346</u>: pH 8.5

For <u>DSM 109470</u>, <u>DSM 109471</u>: PH 5.9

For <u>DSM 109757</u>: cover agar with liquid medium 830

For <u>DSM 110366</u>: Only 20 mg/L of medium 830