

853b. FUSIBACTER BIZERTENSIS MEDIUM

NH ₄ Cl	1.00	g
K ₂ HPO ₄	0.30	g
KH ₂ PO ₄	0.30	g
MgCl ₂ x 6 H ₂ O	0.50	g
CaCl ₂ x 2 H ₂ O	0.10	g
KCl	0.10	g
NaCl	3.00	g
Yeast extract (OXOID)	1.00	g
Trypticase peptone (BD BBL)	1.00	g
Trace element solution (see medium 141)	10.00	ml
Na-resazurin solution (0.1% w/v)	0.50	ml
Na ₂ S ₂ O ₃ x 5 H ₂ O	3.16	g
L-Cysteine-HCl x H ₂ O	0.50	g
Na ₂ CO ₃	1.50	g
D-Glucose	3.60	g
Na ₂ S x 9 H ₂ O	0.30	g
Distilled water	1000.00	ml

Dissolve ingredients, except thiosulfate, cysteine, carbonate, glucose and sulfide, then sparge medium for 30 - 45 min with 80% N₂ and 20% CO₂ gas mixture to make it anoxic. Add thiosulfate and cysteine, then adjust pH to 7.0 and dispense medium under 80% N₂ and 20% CO₂ gas atmosphere into anoxic Hungate-type tubes or serum vials and autoclave. After autoclaving, add glucose and sulfide from sterile anoxic stock solutions prepared under 100% N₂ gas and carbonate from a sterile anoxic stock solution prepared under 80% N₂ and 20% CO₂ gas atmosphere. Adjust pH of complete medium to 7.2 – 7.4, if necessary.