

**898a. ANAEROBACILLUS MEDIUM**

KH <sub>2</sub> PO <sub>4</sub>	0.20	g
MgCl <sub>2</sub> x 6 H <sub>2</sub> O	0.10	g
KCl	0.20	g
NH <sub>4</sub> Cl	1.00	g
NaCl	15.00	g
Trace element solution SL-10 (see medium 320)	1.00	ml
Na-resazurin solution (0.1% w/v)	0.50	ml
Na <sub>2</sub> CO <sub>3</sub>	2.76	g
NaHCO <sub>3</sub>	10.00	g
Yeast extract	0.50	g
D-Glucose	5.00	g
Vitamin solution (see medium 141)	10.00	ml
Na <sub>2</sub> S x 9 H <sub>2</sub> O	0.50	g
Distilled water	1000.00	ml

Dissolve all ingredients except carbonates, yeast extract, glucose, vitamins and sulfide, then sparge medium with 100% N<sub>2</sub> gas for 30 – 45 min to make it anoxic. Add carbonate and hydrogencarbonate, adjust pH to 8.0 - 8.5, then dispense under 100% N<sub>2</sub> gas atmosphere into anoxic Hungate-type tubes or serum vials and autoclave. Add yeast extract, glucose, vitamins and sulfide from sterile anoxic stock solutions prepared under 100% N<sub>2</sub> gas. Vitamins should be sterilized by filtration. Adjust pH of complete medium to 8.5 – 9.0.

For [DSM 18345](#) replace glucose with 2.50 g/l D-mannitol as substrate and adjust pH of complete medium to 9.5 – 9.7.

For [DSM 18979](#) omit glucose and increase amount of yeast extract to 2.00 g/l.